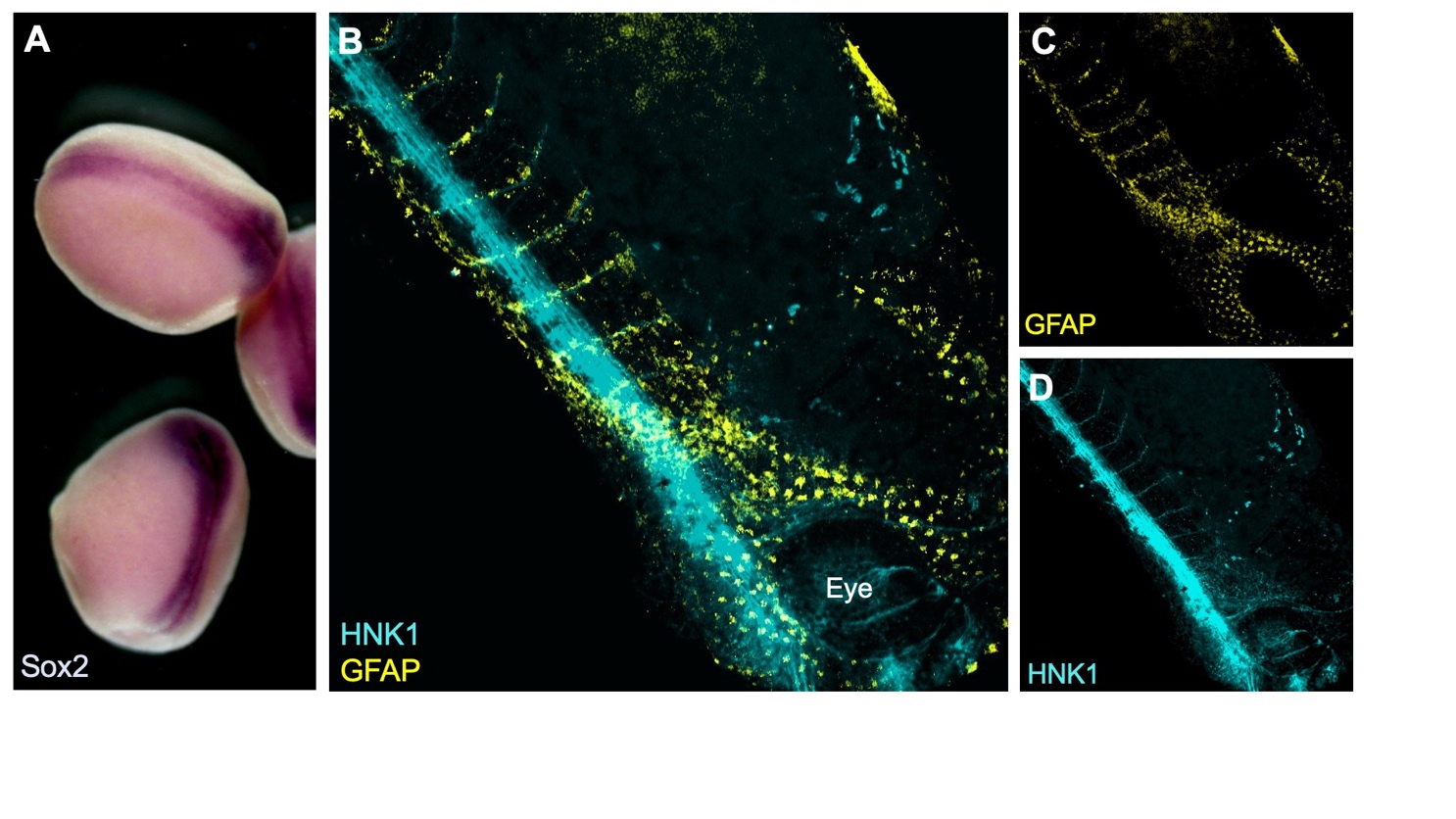
**Visualising neural crest induction, migration and differentiation in *Xenopus***

Throughout my undergraduate studies I have become increasingly captivated by the intricacy and elegance of animal development. Especially interesting to me is how processes such as morphogenesis, tissue patterning or cell migration, which can appear incomprehensible, emerge from relatively simple interactions at the molecular level. Observing how a single cell transforms into a complex organism is a unique and thought-provoking experience. I am grateful to the BSDB for the studentship allowing me to explore this field. I would strongly recommend the Gurdon studentship to anyone interested in topics ranging from evolution, epigenetics, cell signalling, cancer, to stem cells and regeneration – all of these processes may be elucidated by taking a developmental point of view.

During the summer studentship I have learnt to work with *Xenopus laevis* embryos and keep track of their development. The aim of my project was to optimise techniques for visualising the neural crest with other tissues relevant to its induction, migration and differentiation. To do this I used two well established techniques in developmental biology: *in situ* hybridisation and immunofluorescence (see Figure 1).



**Figure 1:** A) Colorimetric *in situ* hybridisation with neural plate marker Sox2 in late neurula stage *Xenopus* embryos. B-D) Whole-mount fluorescent immunostaining for neuronal marker HNK1 and glial marker GFAP in a tailbud (Neiuwkoop and Faber stage 31) embryo.

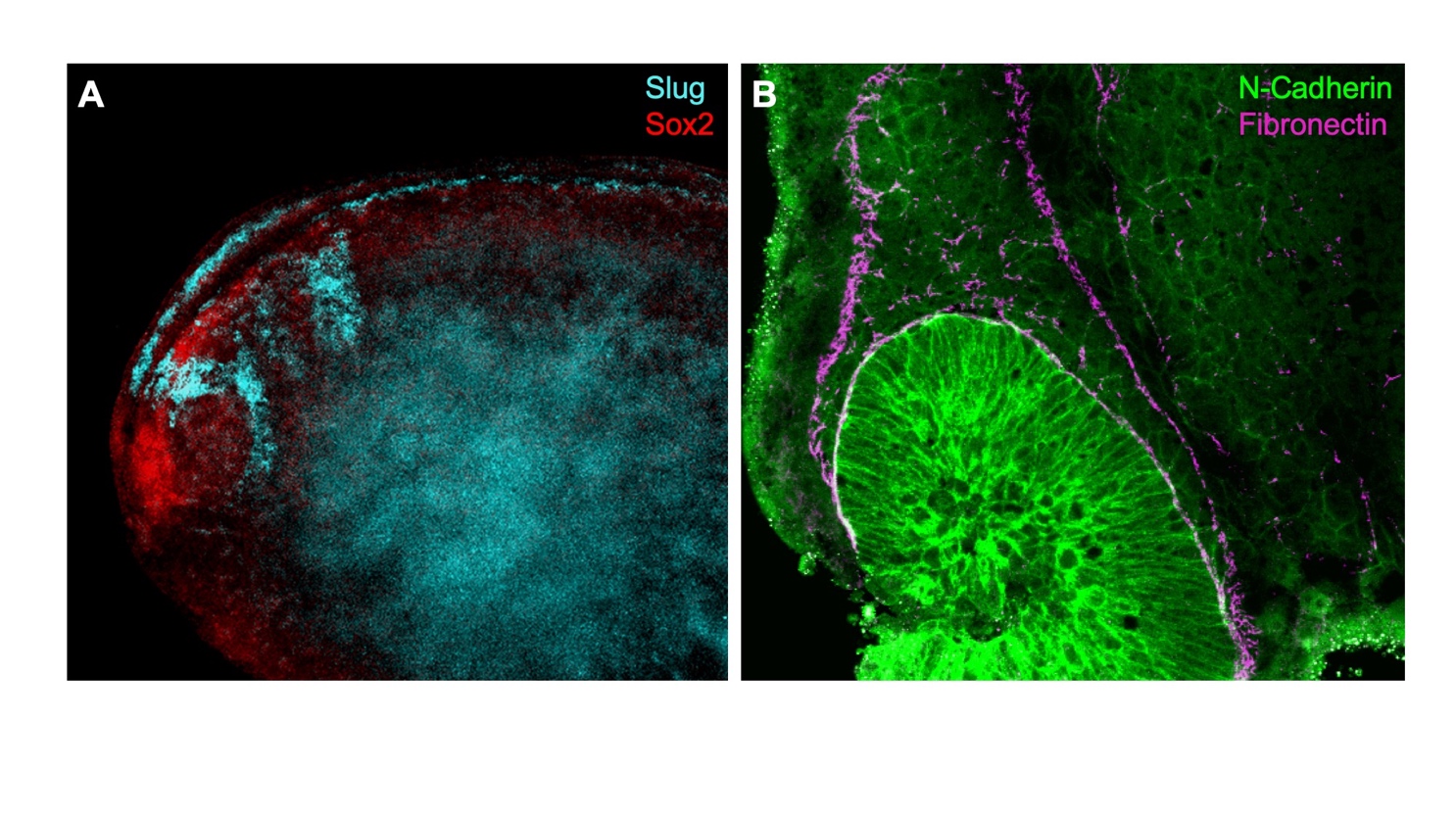
**Why study neural crest cells?**

The neural crest is a fascinating population of cells unique to vertebrates, which is induced at the neural plate border and following neurulation delaminates and migrates away from the neural tube. A subset of these cells migrates into head regions where it gives rise to a variety of tissues including bones, cartilage, as well as neurons. This cranial neural crest also migrates into the branchial (or pharyngeal) arches and contributes in a major part to the craniofacial skeleton. This has undergone major changes during vertebrate evolution. For instance, some of the jaw bones present in our common ancestors with reptiles have given rise to the middle ear bones of mammals (Santagati & Rijli, 2003). A possible explanation for this versatility of the neural crest is that its cells retain multipotency for longer than the three embryonic germ layers, prompting some to consider it a ‘fourth germ layer’ (Simoes-Costa & Bronner, 2015).

The diversity of neural crest-derived cells, which also includes melanocytes, Schwann cells, meninges or the cornea, makes it an important model of differentiation with a potential for therapeutic applications. It is also a great model for studying cell migration (Szabó & Mayor, 2018) and by proxy epithelial to mesenchymal transition in cancer metastasis. Recent work even attributed the ability to remove cellular debris from early neural tube to migrating neural crest cells, potentially through a macrophage-like mechanism (Zhu, et al., 2019).

**Going further with established techniques**

I have been testing and optimising techniques to visualise proteins and gene expression that may help elucidate how the neural crest gets induced and how it migrates through the embryo. For example, I have image the neural crest with nearby mesoderm, which is required during induction, using a combination of fluorescent *in situ* hybridisation (FISH) and immunostaining. In order to visualise expression of two different genes within the embryo a double *in situ* hybridisation (ISH) can be used. This is especially important in *Xenopus*, as no antibody is known to efficiently and exclusively label the neural crest. Although colorimetric ISH is easier and does not require clearing of the tissues, it doesn’t enable exploring the 3D structure by imaging the whole embryo at once. I have used confocal and multiphoton microscopy to analyse the embryos (see Figure 2). Unfortunately, it seems that using two RNA probes at the same time reduces signal intensity and although I have tried to optimise the signal amplification reaction and bleaching to neutralise endogenous peroxidase activity, I have not been able to reduce the background.



**Figure 2:** A) Fluorescent *in situ* hybridisation using RNA probes for Slug (neural crest) and Sox2 (neural plate) in a whole-mount stage 22 *Xenopus* embryo. B) Dissected embryo immunostaining shows N-Cadherin expression in migratory neural crest. Fibronectin outlines the streams of cranial neural crest.

One possibility is to delimit the neural crest using an antibody for fibronectin, an extracellular matrix component that encourages neural crest migration. Using double immunostaining (Figure 2B) I was able to confirm migratory neural crest cells express N-Cadherin near the optic vesicle as fibronectin outlines the neural crest streams (Scarpa, et al., 2015). Towards the end of my summer project, I have also manipulated cell contractility using drug treatments to observe changes in neural crest cell behaviour using several antibodies.

I thank Prof Roberto Mayor for supervising this summer project and Dr Adam Shellard for teaching me methods used in the lab. I have learnt a lot about how research is done and presented my results at a lab meeting in the final week and used this experience to transition from undergraduate study into my PhD.

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